

## Using peptides to study and modulate protein interactions for therapeutic purposes

Assaf Friedler

Institute of Chemistry, the Hebrew University of Jerusalem, Givat Ram, Jerusalem 91904, Israel  
[assaf.friedler@mail.huji.ac.il](mailto:assaf.friedler@mail.huji.ac.il)

### Abstract

Our group is interested in using peptides for the quantitative biophysical and structural analysis of protein-protein interactions (PPI) in health and disease. Based on this, we develop lead compounds that modulate PPI for therapeutic purposes. Peptides serve as major tools both for studying PPI and for modulating them (by inhibition or activation). In my talk I will present an overview of our research, focusing on the following examples:

1. Using peptides to modulate the oligomeric state of proteins<sup>1,2</sup>. Many disease-related proteins are in equilibrium between active and inactive oligomeric states. Specific binding of peptides to one of the oligomeric states of such protein should result in stabilization of this state and consequently in shift of the oligomerization equilibrium towards it. We term peptides with such activity as “shiftides”, and they can be utilized therapeutically in two manners: (i) inhibiting a protein by shiftides that bind preferentially to its inactive oligomeric state and stabilize it; (ii) activating a protein by shiftides that bind preferentially to the active oligomeric state and stabilize it. Examples for shiftides developed in our lab will be described for the HIV-1 integrase<sup>2,3</sup>

2. Development of new synthetic methods for peptide modifications. We developed a new approach for peptide cyclization during solid phase synthesis under highly acidic conditions<sup>4</sup>. Our approach involves simultaneous in situ deprotection, cyclization and TFA cleavage of the peptide, which is achieved by forming an amide bond between a lysine side chain and a succinic acid linker at the peptide N-terminus. The reaction proceeds via a highly active succinimide intermediate, which was isolated and characterized. Our new methodology is applicable for the formation of macrocycles in solid phase synthesis of peptides and organic molecules. We also developed a new general N-acetylation method for solid phase synthesis<sup>5</sup>. Malonic acid is used as precursor and the reaction proceeds by in situ formation of a reactive ketene intermediate at room temperature.

3. Intrinsically disordered proteins as drug targets: About one third of the genome encodes for intrinsically disordered proteins (IDPs) or disordered regions in proteins (IDRs). These lack stable tertiary structures and are composed of a large ensemble of extended and flexible conformations interchanging dynamically. IDPs are involved in many human diseases, making them attractive targets for drug design. However, more than 90% of current drug targets are enzymes or receptors and IDPs still cannot be targeted due to the lack of specific binding pockets for small molecules. Our research focuses on how intrinsic protein disorder regulates protein activity with the ultimate goal of setting IDPs and IDRs as therapeutic targets. In my talk I will describe our studies regarding several IDRs including: (1) the pro-apoptotic ASPP2 protein<sup>6,7</sup>; (2) The HIV-1 Rev protein<sup>8</sup>; (3) the centrosomal STIL protein, which is upregulated in cancer<sup>9</sup>.

### References

1. Gabizon, R. and Friedler A. (2014) *Front Chem.*;2:9
2. Hayouka, Z., et al., (2007) *Proc Natl Acad Sci U S A.*, 104(20): p. 8316-21.
3. Hayouka, Z., et al., (2010) *Bioorg Med Chem.*, 18(23): p. 8388-95.
4. Chandra K, et al.(2014; *Angew Chem Int Ed Eng*, 53(36):9450-5
5. Chandra K, et al. (2014) *Org Biomol Chem.* 12(12):1879-84
6. Rotem, S. et al. (2008) *J Biol Chem* 283, 18990-18999,
7. Rotem-Bamberger, S. et al. (2013) *PLoS One* 8, e58470,
8. Faust, O. et al. (2014) *Chem Comm* 50(74):10797-800
9. Amartely, H. et al. (2014) *Chem Comm*, 50(40):5245-7.